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RESEARCH ARTICLE

The impact of malaria coinfection on Ebola virus disease outcomes: A systematic review and meta-analysis

Hannah M. Edwards *, Helen Counihan, Craig Bonnington, Jane Achan, Prudence Hamade, James K. Tibenderana

Malaria Consortium, London, United Kingdom

* hmedwards88@gmail.com

Abstract

Introduction

Viral outbreaks present a particular challenge in countries in Africa where there is already a high incidence of other infectious diseases, including malaria which can alter immune responses to secondary infection. Ebola virus disease (EVD) is one such problem; understanding how *Plasmodium spp*. and Ebolavirus (EBOV) interact is important for future outbreaks.

Methods

We conducted a systematic review in PubMed and Web of Science to find peer-reviewed papers with primary data literature to determine 1) prevalence of EBOV/*Plasmodium spp.* coinfection, 2) effect of EBOV/*Plasmodium spp.* coinfection on EVD pathology and the immune response, 3) impact of EBOV/*Plasmodium spp.* coinfection on the outcome of EVD-related mortality. Random effects meta-analyses were conducted with the R package meta to produce overall proportion and effect estimates as well as measure between-study heterogeneity.

Results

From 322 peer-reviewed papers, 17 were included in the qualitative review and nine were included in a meta-analysis. Prevalence of coinfection was between 19% and 72%. One study reported significantly lower coagulatory response biomarkers in coinfected cases but no difference in inflammatory markers. Case fatality rates were similar between EBOV(+)/PI (+) and EBOV(+)/PI(-) cases (62.8%, 95% CI 49.3–74.6 and 56.7%, 95% CI 53.2–60.1, respectively), and there was no significant difference in risk of mortality (RR 1.09, 95% CI 0.90–1.31) although heterogeneity between studies was high. One in vivo mouse model laboratory study found no difference in mortality by infection status, but another found prior acute *Plasmodium yoeli* infection was protective against morbidity and mortality via the IFN-y signalling pathway.

Conclusion

The literature was inconclusive; studies varied widely and there was little attempt to adjust for confounding variables. Laboratory studies may present the best option to answer how pathogens interact within the body but improvement in data collection and analysis and in diagnostic methods would aid patient studies in the future.

Introduction

Viral epidemics have been recognised as an increasing global public health threat. As has been observed during the COVID-19, SARS and MERS outbreaks, our increasingly connected world allows rapid international spread of viruses like never before [1-5]. Spread of such pathogens to the African continent is of particular concern not only because of weak surveillance and health response systems, but also because of the high burden of other febrile infectious diseases, leading to problems with accurate diagnoses and patients afflicted with more than one infectious agent at a time [6]. Coinfections may alter immune responses and the expected course of infection, morbidity and mortality, perhaps requiring a different approach to case management among such individuals [7, 8].

Since malaria is highly prevalent across sub-Saharan Africa, it is reasonable to expect the prevalence of viral-Plasmodium spp. co-infection to be high in these settings. Understanding how Plasmodium spp. (herein Plasmodium) infection may alter responses to viral infection, and vice versa, is thus important for their treatment and control. Insights into this may be inferred from Plasmodium's interaction with other established viral infections. Outbreaks of Ebola virus disease (EVD) occur sporadically in countries in Africa where there is already a high burden of malaria. Historically, the majority of EVD outbreaks have been relatively small with case numbers typically less than 100 people [6]. This, combined with the fact that the epidemics can progress very rapidly, occur in remote areas with dispersed populations, and that the affected countries already have weak disease surveillance systems and laboratory diagnostic facilities, means opportunities to investigate the epidemiology of EVD and other comorbidities were limited. The EVD outbreak of 2014 was massive in comparison, with 28,616 suspected, probable and confirmed cases and 11,310 deaths across the three majorly affected countries: Guinea, Liberia and Sierra Leone [6]. In 2013, the incidence of confirmed malaria cases in these three countries was 18, 280 and 279 per 1,000 population in Guinea, Liberia and Sierra Leone, respectively (though, this seems low for Guinea and was reported as 148/1,000 population in fiscal year 2014) [9, 10]. The huge local, national and international effort to control this EVD outbreak led to the collection of large amounts of data from health facilities and has allowed much greater investigation into the clinical and public health effects of Ebolavirus (EBOV) infection [11-13].

Plasmodium has previously been implicated in modulating the immune response to a secondary infection through activation of pro-inflammatory pathways [14–16]. In the case of respiratory viruses, this has a beneficial effect in preventing development of pneumonia in children, whereas it leads to uncontrolled bacterial growth and increased mortality with coincident *Salmonella enterica* infection [17, 18]. *Plasmodium spp.* infection in HIV infected individuals leads to an increase in HIV viral load and could increase risk of HIV transmission and accelerate disease progression [19, 20].

EBOV and *Plasmodium* could interact in several different ways that would be of public health importance if EVD-related morbidity and/or mortality were reduced or increased.

Despite the increase in data available since the West African EVD outbreak, to our knowledge, no one has attempted to synthesize available information on EBOV and *Plasmodium* coinfection and the overall effect on disease outcomes. There has been increasing threat of Ebola viral outbreaks in recent years yet still little is known about how co-morbidities such as malaria infection affect the Ebola immune response and disease outcomes. This, along with the advent of the Ebola vaccine, rVSV-ZEBOV-GP, means it is important to know how these infections may interact to optimise control and case management in the future [21]. With this in mind we set out to review the literature to answer: *How does Plasmodium co-infection affect the immune response, clinical profile and/or clinical outcome to Ebola virus infection*? We present a systematic review and meta-analysis of the literature to determine: 1) the prevalence of *Plasmodium* coinfection on EVD pathology and the immune response to EBOV infection, and 3) the impact of EBOV/*Plasmodium* coinfection on the outcome of EVD-related mortality. Finally, we discuss what gaps, if any, remain in the literature and what the implications are for case management and research in future EVD and other viral outbreaks.

Methods

Search strategy

The literature search was conducted with a final update on 15th October 2020 by the lead study author in PubMed and Web of Science to look for published peer-reviewed papers with primary data on i) prevalence or seroprevalence of *Plasmodium* infection among individuals with current EBOV infection (cases of EVD) or with evidence of past EBOV exposure (indicated by seroprevalence), and/or ii) quantitative or qualitative data on the impact of EVD-related pathology and/or immune response, and/or mortality. To get a complete overview of the evidence we included for review epidemiological studies of infection in humans, clinical case reports and experimental laboratory studies using ex vivo and in vivo infection models. The search was conducted with the following search terms: (Ebola[Title/Abstract]) AND (malaria* [Title/Abstract] OR Plasmodium[Title/Abstract]). While this search strategy was very general, the number of returned articles was deemed feasible to screen without further narrowing of the search terms, thus ensuring all relevant papers could be identified. An initial screen of titles and abstracts excluded irrelevant papers based on being either: review papers, comments/editorials/correspondence, unrelated to EBOV or Plasmodium infection, research on vaccines or drug modes of action, not in the English language. Remaining papers were reviewed in their entirety and papers were included if they met the following criteria: 1) primary peer-reviewed research, 2) either i) epidemiological studies or genomic sequencing studies with explicit measurement of *Plasmodium* and EBOV co-infection prevalence and/or any measure of impact on EVD pathology/immune response/mortality, or ii) clinical case studies with disease pathology/ immune response measures of co-infected patients, or iii) laboratory studies with ex vivo and/ or in vivo models of EBOV and Plasmodium coinfection and a measure of effect on disease pathology and/or immune system modulation and/or mortality. Study of any Plasmodium or EBOV species and within any country and any date were included. In a subsequent meta-analysis, only epidemiological studies with prevalence of coinfection and CFR by malaria infection status and/or effect estimate data on the outcome of EVD-related mortality from countries in Africa were included. We did not include grey literature.

Data extraction

All papers were included for qualitative review of methods and key findings. Observational cohort studies were also included for quantitative review in a meta-analysis. For the meta-

analysis, data were extracted on the following variables: 1) study first author and publication details, 2) study design; 3) location and study period; 4) details of study population, including population size, mean age and sex breakdown; 5) source of data; 6) EVD-related data including method of diagnosis, EBOV species, number of EBOV positive (EBOV(+)) cases; 7) malaria-related data including method of diagnosis, Plasmodium species, number of sample that received a malaria diagnostic test, number of positive tests; 8) Mortality data including crude EVD case fatality rate (CFR), CFR of coinfected patients; 9) Effect estimates (e.g. risk ratios (RR), odds ratios (OR) and hazard ratios (HR)) for the effect of *Plasmodium* coinfection on EVD-related mortality; 10) other variables with a significant impact on mortality (potential confounders).

Extracted data were independently verified (i.e. double verification) by another member of the study team. Where necessary, study authors were contacted to verify study details. The authors conducted this meta-analysis in concordance with PRISMA standards of quality for reporting meta-analyses and the guidelines for Meta-Analyses and Systematic Reviews of Observational Studies [22, 23].

Statistical analysis

We used a random-effects meta-analysis of proportions approach to quantify i) the prevalence of *Plasmodium* coinfection among EVD cases (EBOV(+)/Pl.(+)), ii) crude CFR of EVD cases, iii) CFR of EBOV(+)/Pl.(+) cases and iv) CFR of EBOV(+)/Pl.(-) cases. This was conducted using the metaprop function from the meta package in R [24, 25]. Metaprop allows specific analysis of binomial data and the computation of exact binomial and score test-based confidence intervals [26]. Metaprop was run with a random-effects model to account for heterogeneity between studies in terms of study population, differences in exposure and infection and diagnostic procedures. Random effects models assume the observed studies represent a distribution of possible effects and can incorporate both within-study variance and between-study heterogeneity. To conduct a meta-analysis with individual study weights, we used the inverse variance method of pooling with logit transformation and Clopper-Pearson 95% confidence intervals (CI). The prediction interval was calculated to give a range within which measurements of the same effect size would be expected to fall in future studies. Sub-group analyses were conducted where necessary and as described in the results (i.e. including Zaire EBOV (ZEBOV) species infections only).

For effect estimates related to the impact of *Plasmodium* infection on EVD mortality, where available, numbers of exposed (EBOV(+)/Pl.(+)) and unexposed (EBOV(+)/Pl.(-)) and with and without the outcome of death were used to calculate unadjusted RR and used in a random effects meta-analysis of RR using the metabin function within the meta package which allows meta-analysis of studies with comparison of two groups with binary outcomes [24]. Finally, any adjusted RRs reported in included studies were included in a meta-analysis using the metagen function which performs fixed and random effects meta-analysis based on estimates and their standard errors using inverse variance weighting for pooling [27].

Heterogeneity between studies was quantified for each proportion and effect estimate measure using the I². Chi² and Tau-squared (τ^2) statistics. I² is a measure of the percentage of variability in the effect sizes that is not caused by sampling error, where 25% is considered low heterogeneity, 50% moderate heterogeneity and 75% substantial heterogeneity [28, 29]. Although I² is not sensitive to the number of studies, it is sensitive to the precision of the included studies. Tau² measures the between-study variance within a random effects model and is insensitive to the number and precision of included studies. The square-root of Tau² is the estimated standard deviation of the underlying effects across all the studies [30]. In all models between-study variance was estimated using the Hartung-Knapp-Sidik-Jonkman (HKSJ) method, which is considered to be superior to the standard DerSimonion-Laird method in estimating variance of the pooled effect, particularly when the number of studies is small [31].

Assessment of study quality and bias

Due to the small number of studies ultimately included in the effect size meta-analysis, publication bias via assessment of funnel plot asymmetry could not be assessed since the power to distinguish true asymmetry from chance is limited when the number of studies is <10 and is, therefore, not recommended [32]. However, studies included in the meta-analysis were assessed for quality using the Newcastle-Ottawa Scale, developed to assess the quality of nonrandomised studies incorporated into meta-analyses [33]. The scale uses a 'star system' to judge publications on three key areas: i) selection of study cohort(s), including representativeness of the exposed cohort and measure of, ii) comparability of cohorts based on whether important confounding factors were adjusted for in analyses, and iii) ascertainment of the outcome (for cohort studies) of interest including how it was assessed and whether rates of follow-up were sufficient to avoid potential study bias. In line with previous meta-analyses, included studies were assessed to be good, fair or poor based on the number of stars awarded within each category: "good" meant 3 or 4 stars in selection, 1 or 2 stars in comparability, and 2 or 3 stars in outcomes; fair meant 2 stars in selection, 1 or 2 stars in comparability, and 2 or 3 stars in outcomes; and "poor" meant 0 or 1 star(s) in selection, or 0 stars in comparability, or 0 or 1 star(s) in outcomes. Thus, studies had to achieve at least one star in each section to be deemed good or fair.

Results

Search results

Following removal of duplicates, the initial search returned 322 results which had titles and abstracts screened for eligibility (Fig 1). Following the initial screen, 65 results were reviewed in their entirety and a final 17 papers were included for qualitative review as they had a measure of EBOV and *Plasmodium* coinfection prevalence and/or impact on disease outcome according to our inclusion criteria (Table 1). An additional paper was retained since it presented additional demographic information on the same cohort of patients as another which presented coinfection outcome data [34, 35]. We noted potential overlap of the study population between some studies which were confirmed with study authors. Since the studies had different sample sizes, methods and results, all were retained for review and the overlap of populations clearly marked in Table 1. Overlapping studies were excluded in the quantitative meta-analysis so that the study with the largest study population was included in each instance.

The final set of papers qualitatively reviewed thus included 14 observational studies (13 cohort studies and one cross-sectional study), two laboratory studies and one clinical case report. A final set of nine cohort studies were included in the quantitative meta-analysis based on having a unique cohort of patients with similar patient characteristics. Studies were excluded if drawn from the same sampled cohort as another more up-to-date or larger study (n = 3), or if they sampled only children (n = 1).

Cohort studies. Nine retrospective cohort studies [34, 36-43] and four prospective cohort studies [44-47] were identified, all with health-facility based cohorts of suspect and/or confirmed EVD cases from Ebola treatment units (ETUs) in Liberia (n = 3), Sierra Leone (n = 2), DRC (n = 2), Guinea (n = 4), and with sites in both Liberia and Sierra Leone (n = 2) although



Fig 1. PRISMA flow diagram for study selection.



these only included malaria testing in Sierra Leone. Due to overlap of study populations, however, these 13 studies drew from nine unique populations (as indicated in Table 1).

Sample size of the study cohorts varied from 52 to 2741 and EVD cases from 52 to 1512. Twelve of the studies included all age groups in their cohort, while one included only children aged <18 years. Percentage of males among EVD(+) cohorts was mostly between 44 and 52% (n = 5), although ranged from 13.5% to 63%.

EVD was diagnosed by real-time PCR (RT-PCR) in eleven cohort studies including seven studies [34, 36, 38, 40, 42, 46, 47] which used quantitative RT-PCR (qRT-PCR) through which cycle threshold (C_t) was analysed as a proxy for viral load. Cycle threshold is defined as the cycle number of the assay at which the fluorescence exceeds a certain threshold, with lower Ct values indicative of higher viral load and vice versa [48]. The remaining two studies utilised a metagenomics (mNGS) approach to identify EBOV RNA in blood and plasma samples from EVD patients. One of these used a combination of RT-PCR test data from both DRC and US-based laboratories and mNGS results to determine whether patients were confirmed (at least two positive tests) or probable (one positive test) EVD cases. Although rarely explicitly stated, from the outbreak date and location, most of the observational studies were inferred to be

Observation	nal studies				-		-				
Population Id	Study #	Publication	Study design	Study location and study period	Study Population	Source(s) of exposure and outcome data	EVD data	Malaria data	Outcome data	Effect estimate (unad), and adj.)	Included in meta-analysis?
-		Barry et al. Med Mal Infect. 2014; 44(11–12):491–4	Prospective health facility-based cohort	Conakry, Guinea Mar-Àug 2014	Patients with confirmed EVD (by KTP-QC) admitted to EVD (by KTP-QC) admitted to a single ETU (N = 90) Demographics - Male 63% (n = 57) - Male 63% (n = 57) - Male 63% (n = 57) - Male 64% (n = 54) - Male 70% (n = 54) - Days from symptom onset - Days from symptom onset to hospitalisation: 6 ± 3 days	Data collected prospectively by trained medical staff	EVD(+) cases: 90 Bagnostic: KT-PCR EBOV species: Zaire Viral load: N/A	Plasmodium infection among RYD(+) asses: Tested = 90 Positive = $65/90$ (72%) Diagnostic: not reported Species: unspecified Parasite load: N/A	Crude CFR: 39/90, 44% (95% CI 33-54) CFR of coinferted cases: N/ A	N/N	No-CFR by malaria status noi reported, just prevalence of målaria co- infection
0	0	Kraiz et al. PLoS One. 2015: 10(6): e0129333	Prospective health facility-based cohort	lsino, DRC Aug-Slov 2012	Probable & confirmed EVD cases (N = 23) ether: cases (N = 23) ether: in stor (N = 18) in stor (N = 14) perographics community (N = 34) Demographics or Mate 23,1% (or = 12), incl. (n = 7),14.7% community cases (P = 7), 14.7% community (n = 7),14.7% community (n = 7),14.7% community cases (P = 7), 14.7% community cases (P = 7), 14.7% community or expresented in overrepresented in community and ETU cases community and ETU cases within 3 days of symptom onset	Demographic and clinical data obtained from routine data collected by ETU staff collected by ETU staff	EVD(+) cases: 52 EVD(+) cases: 52 EVD(+) • 34 community cases (16 • 34 community cases (16 Diagnostic: PCR or lgM-EILSAIEOV species: BDBVViral load: N/A	Plasmodium infection mong RYD(+) cases: Tested = 9 Positive = 4/9 (44.4%, three positive RPTs and one treated as presumptive malaria (unspecified if Pf only or Pf/pn) Species: unspecified Parasite load: N/A	Crude CFR: 28/52, 53.8% Including: CFR of community cases: 55.9% (community cases: 55.9% (93.8%) probable cases, 22.2% confirmed cases, 22.2% confirmed cases, 20.0% (all confirmed cases) 0.0% (all confirmed cases) Concomitant malaria did not influence CFR (CFR and int, raw numbers not provided)	N/N	Yes— prevalence and cFR by malaria status reported status reported
m	m	Gignoux et al. New Engl J Med. 2016. 73374(1):23 32	Retrospective health facility- based cohort	Foya. Liberia Jun-Oct 2014	Confirmed EVD patients admitted to single ETU (N = 381) Demographics Demographics - Alac $4.3%$ ($n = 184$) - Alac $4.3%$ ($n = 184$) - Alac $4.2%-$ Alac $4.2% -$	Data compiled by staff epidemiologist from clinical fries and lab results	EVD(+) cases: 381 BBOY species: Zaire BBOY species: Zaire Viral load, median PCR C1 at admission: 194 (IQR 171–22.8)	Plasmodium infection among EV-16, saxes: 337, 19, 3% 337, 19, 3% Dagnestic: BinaxNOW RDT Species: unspecified although RDT detects all four primary species Parasite load: N/A	Crude CFR, 202/328, 61.6%, (53 cases with missing outcome data) AJ56, 60.7%, CFR of coinfected patients: AJ56, 60.7%, 60.3%, 60.3%	Increased risk of death with Plasmotion: Oundecourd. Justic RR = not reportedaRR = 1.23, 95% CI 0.53-2.86, p = 0.63 (adjusted for antimatical prescription, age goup, actimatical prescription, age goup, sympton onset to admission, trencipt of treatment centre on day of admission. Full data has used on some and the admission and the admission of the admission and the admission. Antimatical treatment: ASAQ treatment associated with lower risk of death han A-L, all 80, 695, 95% CI 0.54- 0.89 and a symptom onset to admission, JV fluids and a inpatient darg treatment - ASAQ was productive admission, JV fluids and a inpatient darg for analysis showed malina infection may inhibit the response to admission, JV fluids and a inpatient darg for analysis showed malina infection may inhibit the response to admission, JV fluids and a inpatient darg for analysis showed malina infection may inhibit and a inpatient darg for a admission, JV fluids and a inpatient darg for a admission, JV darg for admission, JV fluids and a inpatient darg for a admission, JV darg for admission, JV darg for admission, JV darg darg for admission, JV darg for darg for admission, JV darg darg for darg f	Yes— CFR by mainta FR by mainta status reported and effect estimate
	4	De Wit et al. Emerg Infect Dis. 2016. 22(2):323-6.	Retrospective cohort/Diagnostic feasibility	Monrovia, Liberia 2015 2015 2015	Samples from suspect EV D patients submitted to single diagonations in the single (N = 1,056) Demographics not reported	Data from CDC-NIH ELWA diagnostic laboratory	EVD(+) 306/1058 Dagnostic RT-PCR Species Zaire Viral load: N/A	$\begin{array}{l} Plasmodium infection\\ overall sample:\\ rested = 1038\\ Positive = 200\\ Positive = 200\\ Positive = 200\\ Plasmodium Coinfection:\\ 47/306 (15.4%)\\ Diagnostic: qRT-PCR\\ (Ci \leq 30 considered error operative used to make operative used to make operative used to make operative used to make positive used to make analysis analysis (Plasmodium included in analysis (Plasmodium included in Paramodium included in Paramodium included in Plasmodium included in Plas$	N/A	N/A	No-overlap of study population with Rosenke et al. and no CFR reported
											(Continued)

Table 1. Details of studies included in the qualitative and quantitative reviews.

Yes- prevalence and cFR by malaria status sported and effect estimate estimate	Yes— Prevalence and CFR by malaria status reported and effect estimate	No.—overlap of study population with Kerber et al.	(Continued)
RRs reported as effect on survival. <i>Plasmidium</i> coinfictum associated with increased survival. Unadj. RR = not increased survival. (21, 11-14, $\beta = 0.004$) arRs 10, 21, 11-14, $\beta = 0.004$) arRs 10, 21, 11-14, $\beta = 0.004$ <i>Plasmodium</i> burden (Ct < 200) = RR 14, 12-17, $p \sim 0.0001$ Low <i>Plasmodium</i> burden (Ct < 200) = RR 14, 12-17, $p \sim 0.0001$ Low <i>Plasmodium</i> burden (Ct > 200) = RR 12, 038-14, $p = 0.08$ arR adjusted for age, sex and EBOV Ct kevel	Odds of faality ascessed among 1047 EVD(-) patients with complete datasets EVD(-) patients with complete datasets EBOV(-)/Pl(-): reference EBOV(-)/Pl (-): 1.2.5% C(J0-).2; p= 0.3.2 (-): 1.2.5% C(J0-).2; p= 0.3.2 MS for fatality with interaction between <i>Plasmituri</i> infection and age, among 5: 14yr sag group; 5: 14yr sag group; 8: 16yr sag group; 9: 10,11, p= 0.002 0 other age group; adjusted for EBOV Cr value. Other sig variables: EBOV Cr value Other sig variables: EBOV Cr value	V/N	
Crude CFR: 612/1182, 51.8% Cointected CFR2.8(128, 42.2%CFR EBOV(4)/P((-); 54%(P = 0007)CFR by Plasmotiants C1evei: C1 < = 20.529, 172% - C1 > 20.739, 156, 46.7% - C1 > 20.73156, 46.7% - C1 > 20.735, 45.7% - C1 > 20.735, 45.7% - C1 > 20.735, 45.7% - C1 > 20.735, 45.7% - 11004, 10	Crude CFR: 719/1205, 59.7% among hospitalised EVD patients Mortality rate highest among patients aged < 5 years and >74 years (80 and 90% respectively). CFR by infection status: EBOY(+)/P(L(-): 150/249, 60.2% EBOY(+)/P(L(-): 452/798, 56.6% S6.6% EBOY(+)/P(L(-): 435/69, 62.3% EBOY(+)/P(-): EBOY(+)/P(-): EBOY(+)/P(-): 22/54.40.7%	Using RDT data alonc- Dismotium increased patient mortality ($p = 0.053$, ANOVA) Using sequencing data – Using sequencing data – Dismotism experiencing the highest burden of mortality rate of SWS ($n = 6$), higher than patients with low parate burden Mortality increased with <i>Plasmodium</i> sequence depth, but no absoluce depth, but no absoluce BEOV(+)/P(+) ($n = 13$) BEOV(+)/P(+) ($n = 13$) associated with the EBOV(+)/P(+) ($n = 13$) associated innae (FBOV volt markers associated on badditon al effect on the innae (markers) associated with the essociated with the cognitation pathway were increased in patients:	
Plasmodium infection among EVD(+) cases: Tested = 956 Positive = 183/956, 19% positive Drs qRT=PCR. Cf \leq 30 Drs qRT=PCR. cf \leq 30 considered positive Plarasite bad, menn Cf \pm SD: 24.8 \pm 4.0	Plasmodium prevalence overall: 1937/2178 (89%) hospital attendees Confrection prevalence among hospital EVD cases 261/1231 (24%) Disgnostic: BinaxOW Bigmostic: BinaxOW RDT (detects Pf/pan) Disgnostic: BinaxOW RDT (detects Pf/pan) atthough RDT can detect all four major species	Plasmodium infection: RDT results (from patient records) = 40(21, 33% Sequenced reads = 156/186, 89% blood ad plasma samples contained reads mapping to Pf Read depth varied widely (FFKM value as masure of read depth varied from 9) to 3588201)	
EVD(+) cases: 1182 Dagnostic qRT-PCR Specta: Zaire Viral load, mean Ct ± SD: 27.0 ± 4.7	EVD(+) confirmed: 1231/2178 BVD(+) confirmed: 1231/2178 community denths (59%) Dagnostic qRT-PCRSpecies: ZareVral Joad and survived 23.2, 2.42.5.5 H-Osphalised and died 18.1, 1.6.2-2.08 • Community deaths 21.5, 19.4- 24	EBOY infection: Reads mapped to Zaire ebolavirus in all samples	
Demographic and clinical data sourced form patient data forms whom theth of algenosic lab and updated with lab cutcome determined by cross-referencing names on patient data forms with a list of deceased patients at ETU	Laboratory request forms updotaded to BMLab database—cross referenced with Guinean EVD pattent database	Clinical patient records Blood and plasma sumples sequenced using HiSeq 2500 system	
EVD(+) confirmed cases admitted to single ETU (N = 1182) Demographics - Maler 43-3% - Maler 43-3% - Mane 29-38 ± 15.6 (range 0.01-33) (range 0.01-33) - Systar not included in effect size estimates due to low numbers	Suspect EVD cases attending hybridization in the initial control of current (N = 2128) and community deaths (N = 563) with amples submitted to EMLab unt in Garcisádou, Guine a Demographics or Male 47% supect hopfal cases, 57% community deaths 44) suspect hospital cases, 37 (25–55) community deaths (25–55) community deaths	EVD cass diagnosed by European Mobile Laboratory in Guéckédou, Guine including hosptalised putients and deaths in the community N = 202, incl: N = 202, incl: (arrange 15, 23, 11, 01) (arrange 15, 23, 11, 01) (arrange 15, 23, 11, 01) (arrange 15, 23, 11, 01) (arrange 12, 23, 11, 01) (arrange 12, 23, 11, 01) (arrange 12, 23, 11, 01) (arrange 12, 24, 11, 01) (b) (b) (b) (c) (c) (c) (c) (c) (c) (c) (c) (c) (c	1
Monrovia, Libera Aug 2014-Feb 2015 2015	Guinea (cases mosty from Macerna and Guéckédou)	Guinea Guinea	
Retrospective health facility- based cohort	Prospective health facility-based cohort	Metagenomic RNA Deep-sequencing study retrospectively sampled patient cohorts	
Rosenke et al. Clin Infect Dis. 2016. 63(8): 1026–33	Kerber et al. J Infect Dis. 2016 0.15: 214/Suppl 3): S250-S257 3): S250-S257	Carroll et al. msphere: 2017 2 (4):e00325-17	
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Table 1. (Continued)

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Yes— prevalence and CFR by malaria status reported and effect estimate	Yes— prevalence and CFR by malaria status reported	No-overlap of study study Waxman et al, children aged <18 years only	Yes— prevalence and prevalence and status reported and effect estimate estimate (Continued)
OR for fatility: EBOV(+)/Pf(-) = reference EBOV(+)/Pf(-) = 2, p=0.03 (BJ)V(+)/Pf(-) = 3, p=0.03 (BJ)V(+)Pf(-) = 3, p=0.03 (BJ)V(+)Pf(-) = 3, p=0.03 (BJ)V(+) Pf(-) = 3, p=0.03 (F)V(-) Pf(-) = 3, p=0.03 (F)V(-) Pf(-) = 2, p=0.03 (F)V(-) Pf(-) Pf(-) Pf(-) = 2, p=0.03 (F)V(-) Pf(-) Pf(-	CFR significantly assoc. with baseline viremia	CFR significantly associated with age, bleeding, median initial EBOV Ct, ength of hospital stay initial EBOV Ct, <i>Plasmidium</i> parasitentia had no <i>Plasmidium</i> parasitentia had no impact on EVD outcomes in aggregate (p = 0.76) or age-straified analyses (data not shown)	Cox proportional hazatds model for effect on mortality: effect on mortality: EBOV(-)/PL(-) = reference group - EBOV(-)/PL(-) aHR = 6.0, 4.4–8.0, EBOV(-)/PL(-) aHR = 6.0, 4.4–8.0, EBOV(-)/PL(-) aHR = 6.0, 4.4–8.0, P = 0.001 - EBOV(-)/PL(-) aHR = 0.4, 0.2–0.7, p = 0.001 - EBOV(-)/PL(-) aHR = 6.0, 4.4–2.5, p = 0.001 - BOV(-)/PL(-) aHR = 6.0, 4.4–2.5, p = 0.001 - BOV(-)/PL(-) aHR = reference p = 0.001 - aHR = reference group/l(-) aHR = reference group/l(-) by
CFR: Crude = 6/158, 60,8%, EBOV(+)/P(t+) = 26/35, 74.3% EBOV(+)/P(t-) = 53,6%	Crude CFR, 57/97, 58.7% CFR EVD(+)/Pf(+): 12/14, CFR EVD(+)/Pf(-): 36/60, 60.0% (p = 0.12)	Crude CFR 69/122, 56.6%	CFR: Crude = 140/254, 55, 1%, 95% CI 48.8 - 61.3%, EBOV (+)/PL(+) = 66%, 35/ 33 CL (+)/PL(-) = 22%, 105/ 201 28 - duy morthy highest in C38 - duy morthy highest in C38 - duy morthy 95% CI 59-11.4)
Plasmodium infection in overall sample: Tested = 543/566 Positive = 188, 34.6% Plasmodium coinfection among EVD(-) cases: Prevalence = 53(14, 24.1% Za greater can do of malaria infection among EVD(-) than EVD(-). 38% v. 24%. Za greater and so finalaria infection highest in <5 yrs Prevalence of Plasmodium infection highest in <5 yrs Prevalence of Plasmodium infection highest in <5 yrs prevalence of an only Clatters Pf only) Species: unspecified but analy Plagmosted Parasite bads. N/A	Plasmodium infection mong EVD(+) cases: Tested = 74 Positive = 4/74, 18,9% Diagnostic: ONSITE Pf/pan RDT Species: P. falciparum	Malaria testing available from Sterat Leone only. <i>Plasmodium</i> infection among EVD(4) cases: Tested = 6884 Diagnostic: BraaxVOW Diagnostic: BraaxVOM Diagnostic: BraaxVOM distinguishes 4 major distinguishes 4 major species	<i>Plasmodium</i> infection: Deveal sample tested for malaria = 13.68 (10.0%) <i>Prevalance</i> of malaria: BBOV(+) = 53/254, 21%, BBOV(-) = 55/114, 14, 33% Diapostic: Binax/OW Diapostic: Binax/OW Diapostic: Binax/OW Species: unspecified although RDT Species: unspecified datinguished all four major species Parasite load: N/A
EBV diagnosed in 27.9% patients (n = 158/566) (n = 158/566) Species Zaire Species Zaire (Yiral load, mean Cr (range): 22.0 (13.5–37.9); and 39% EVD(+) cases with Cr < 20 with Cr < 20	EVD(+) patients: 97/168 (58%) expendite rel positive Ct ≤ 34 considered positive Viral load, baseline viramia arbitrary units (AU, defined as AU = 2 ^{04-CD}); <-25000 AU (equivalent to Ct 9.55 = 55% ($n = 41$) $\cdot \geq 25,000 = 45\%$ ($n = 33$)	$\begin{split} EVD(+) & cases: 122\\ cases: 122\\ considerationstic eqRT:-DCR, Ct \leq 40\\ considered positiveSpecies Zair load, medication (OQB): survivors 25 (21.3–27.5), died 19.2 (16.9–23.7), p < .001$	EVD diagnosed in 254/1568 patients (19%) Dx mchod: RT-PCR (Ct<40) Species Zinte Viral load: N/A
Routine clinical files	Clinical data from ETU patient records Biological data from Paseur laboratory in Macenta	Clinical data forms digitized into unified database and linked with database and inked with malaria testing malaria testing	Clinical paper forms filled in by medical care staff and uploaded to single database
Suspect EVD patients admitted to a single ETU (GOAL-Mathaska ETU) (N = 566) Demographics: Demographics BEVD4) ag 32.4 yrs, incl. 30.6 EVD4) ag 32.4 yrs, incl. 30.6 EVD4) ag 32.4 yrs, incl. 30.6 eVD4 and everal sample, 4 % Male EVD(4) 4 % Male EVD(4) (4), mean: 4.2 days	Suspect EVD patients admitted to two ETUs ($N = 168$) Cimical follow-up on 77 EVD(+) patients Biodomical analysis on 67 EVD(+) patients and EVD(-) febrie controls febrie controls Controls Febrie controls And Soff = 35 And Soff = 45 Co-9 = 11 (0-9 = 19 Co-9 = 11 (0-9 = 19 Co-9 = 10 Co-9 = 11 (0-9 = 19 Co-9 = 10 Co-9 = 11 (0-9 = 10 Co-9 = 10 Co-	Children <18 years with confirmed EVD (by qRT-PCR) and outcome data at 5 ETU5 (N = 122) Demographics: Male 44% (n = 54) Male 44% (n = 54)	Suspect EVD pattents admitted to three ETUs with test results for both EBOV and Plasmodium infection (N = 1368) Demographics: - Age, median (1QR): overall = 2.9yrs (20-44) EBOV(+)/PL(+) = 30yrs (20- 45) EBOV(+)/PL(+) = 13yrs (4- 53% (n = 715) Male in overall = 20yrs (20-44) coverall = 20yrs (20-44) (4-1)
Port Lko. Sterra Leone Dec 2014-Nov 2015	Matchia and Nearekore, Cainea Nov 2014-jan 2015	Liberia and Sierra Leone 5 ETU's run by IMC Sep 2014-Sep 2015	3 ETUs in Lunsar, Makeni Lunsar, Makeni (run M yu MO, Rerra Lone Doc 2014-Oct 2015 2015
Retrospective health facility- based cohort	Prospective health facility-based cohort	Retrospective health facility- based cohort	Retrospective health facility- based cohort
Harley et al. PLoS Neg Trop Dis. 2007.11(2): e000255 ⁴ e0005356 ⁴	Vernet et al. JCI Insight. 2017. 2 (6):s88864	Smit et al. J Infect Dis 2017. 64 (3):243-249	Waxman et al. 2017. 17(6):654- 660
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Table 1. (Continued)

verlap Vaxman	y malaria reported fiet te	o on the second s		
Noo with W et al.	Yes— Prevale CFR b status and eff estima	No-n messu curren infecti,		2
Covariates significantly associated with mortality = time to ETU opening. Ct value, abnormal bleeding, diarrhoea, dispipaiga and dyspnoea Insignificant difference in mortality by <i>Plasmolium</i> infection status ($p = 0.08$) Some prients assumed exposed to ASAQ MDA showed non-significant decrease in risk of death—RR 0.63, 0.37–1.07, $p = 0.086$	NIA	Due to missing data, analysis of indytali risk factors conducted on 30.2 persons. Positive correlation between geographic distributions (preventine across distributions (prevention coefficient persons) and a second probability of EBOV exposure and Manalum positivity (Spearman rank correlation coefficient persons) and a second probability of current Plasmodium spp. Individual level prior exposure to EPola strongly associated with increased probability of current Plasmodium spp. infection even after accounting for geographic locations and all other individual and population (seef isk probability of current Plasmodium spp. infection even after accounting for geographic locations and all other individual and population (seef isk factors (GR = 1.741 (1.40–2.14) pc-0.0001) EBV analogies was repeared with factor for Plasmodium infection than any other individual trait and second only to thrug in a Lakeland hostita. Associations for all species Plasmodium were qualitatively identical to Pf and Pm care)		BOV RNA levels.
Crude CFR: 244/424, 57.5 EVD(+)/PL(+) CFR: 31/48, 64.6% EVD(+)/PL(-) CFR: 101/ 195, 51.8% EVD(+)/PL(missing) CFR: 112/181, 61.9% Malaria RDT result in survivors vs. died: - 52.5% v 41.4% negative - 53.3% v 45.9% missing	CFR overall: 38/70 patients CFR vorerall: 38/70 patients (57.9% (2 missing outcome data) (CFR by Plasmedium infection status for 25 patients with dinical data available. EBOV(+)/Pf(+) = 10/18, 55.6% PBOV(+)/Pf(+) = 5/7, 71.4% PBOV(+)/Pf(+) = 5/7, 71.4% PBOV(+)/Pf(+) = 5/7, 71.4% CBEOV(+)/Pf(+) = 5/7, 71.4% CBEOV(+)/Pf(+) = 5/7, 71.4% (1 mission difference in clinical datacteristics (e.g. fever, headache, vomiting fever, headache, vomiting fever, headache, vomiting fever, headache, vomiting fever, headache, vomiting fever, headache, vomiting fever, headache, vomiting second on on disease security on nordily but only stangle state only stangle state	CFR. N/A		in terms of RBC number, MA-J eneficial effect
Plasmodium infection data from whole sample: Teaked = 243/424, 57% Positive = 48 (1.1.%) Negative = 195 (4.0.%) Missing = 181 (4.2.7%) Missing = 181 (4.2.7%) From teaked satisst: Positive = 48/243, 19.8% Diagnostic: BinaxNOW Bignostic: BinaxNOW Species: unspecified Parasite load: N/A	Pf data: PF data: PF NA reads NA reads in 21/70 (30.0%) patients, number of reads varied between 1 and 248,600 ers sample, mean 4548 ± SD 29,980 Coinfected amples: 977 Coinfected amples: 977 (24.3%) (24.3%)	Plasmodium infection in whole sample: 22.9% infected with 1 or 22.9% infected with 1 or plasmodium species Plasmodium cointection in seropositive EBOV = 425, 10.2% 10.2		ed but all other mice died. s were observed between groups to coinfected mice observed no L
EVD cases: 424 Diagnostic = qRT-PCR, Ct < 40 Viral load: $C \ge 22 = 123, 37, 5\%$ Ct > 22 = 123, 38% Ct > 22 = 123, 38% Amore available: $Ct \le 22 = 56.6\%$	EVD(+) cases: 2^{2} confirmed (at last 2/3 positive tests) -15 probable (1/3 positive tests) by testkT-PCR in DRC = 31/70 (64 5%)FRT-PCR in US = 21/70 (64 5%)FRT-PCR in US = 21/70 (39%)mMCS reach-mean 8.0%) EBOV reach-mean 8.0%) SD \pm 41,246 (range 1–286,723)	ZEBOV-specific IgG antibody Diagnostic ZEBOV-specific IgG ELISA for seroprevalence Species Zaire	Key results	One mouse infected on dpi-0 surviva No statistically significant difference Repetition of dpi-0 time point with 4
Data obtained from standardised clinical records forms	Samples tested by RT-PCR and NCC repeat RT-PCR and mNCS followed by BBOV- specific capture probe	Random sampling of villages stratified by province—each during field missions from July 2005-Mat from July 2005-Mat of vesson dry cason dry cason dry cason dry cason dry cason dra, and Demographic and Health Survey 2012		traperitoneal inoculation tes), followed jy IA. EBOY on different ne group on each third 1) ministered: MA-EBV 100 e survival, 4 scheduled EBOV inoculation essured
EVD(+) patients admitted to 5 ETUs (N = 424) Demographics • Median age 30yrs (16-44) • 40.6% Male	Suspect EVD patients from Boende.DRC Aug 0014-56p 2014 N = 7014-56p 2014 Including 37 confirmed/ probable EVD cases with demographics 5/37 (1.5.%), Male Mean age 35.4yrs (±16.1)	Fernanent residence aged > 15/synst from 210 rural villags (population < 300) across 9 administrative provinces N = 4.272 enrolled, M = 4.728 male obtained from 4170 incl: 47.3% male median age 49	Methods	Eight groups of 10 mice with in of by (10° parasitized erythrosy intraperional incoulation of A) days post Py inoculation (dp). One group on dp-14 and then o. Ding the job and then o. Appl. (i.e. dp): 2, dp/5. J, dp/5. J, dp/5. Single high dose of MA-EBV a dp) (i.e. dp): 2, dp/5. J dp/5. J Single high dose of MA-EBV a redian lethal dose for mecproy on day 4 post MA- for necropys on day 4 post MA- Outcome: survival over time No immune system markers m
5 ETUs run by IMC, Liberia & Sierra Leone Sep 2014-Sep 2015	DRC	Gabon Survey conducted between July 2005– May 2006	E	infection, time- details: <i>Plasmodium</i> <i>n</i> 17XNL), <i>i</i> EBOV ittenuated EBOV
Retrospective health facility- based cohort	Metagenomic RNA rucospetuky rucospetuky sampled cohort	Cross-sectional	Experimental desig	Murine model of co course studyModel (: . CD1 mice • Model of human infection = Py Study • Model of humar • Model of humar infection = mouses (MA-EBOV)
Garbern et al. Open frorum Infect Dis. 2019. 6 (7).06.250	Li et al. J Clin Microbiol. 2019. 57(9):e00827-19.	Abbate et al. Emerg Infect Dis. 2030. 26(2):229- 237.	ory studies Study	Rosenke et al. 2018. J Infect Dis. 218(suppl 5): \$434.5437.
12	٤ 	14	l laborato Study	# S
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Table 1. (Continued)

	2	Rogers et al. Cell Rep 2020. 30 (12);4041 -4051.e4.	Ex vivo model of coinfection: • Naive human and mouse macophysics • Plasmodium infection: Challenge virth sea from acutely parasitaemic rodents • EDOV infection: non-human primate sear from hasts macaques infected with, <i>p. yamolgi</i> in vivo murine model of coinfection: • Mice BALB(J, p. yamolgi in vivo murine model of coinfection: • Mice BALB(J, p. yamolgi in vivo murine model of coinfection: • Mice BALB(J, p. yamolgi nivo murine model of coinfection: • Mice BALB(J, p. yamolgi nivo murine model of coinfection: • Mice BALB(J, p. yamolgi in vivo murine model of coinfection: • Mice BALB(J, p. yamolgi • Plasmodium infection: Py (& Pc) • Plasmodium infection: Py (& Pc) • BASV infection: Py (Q, PVI-BBOV (oill Mayinga, a variant of Zaire EBOV)	In vivo experiments: 6 groups 10 mice - Day 6 chailenged with 1, 0 or 100 plaque forming inoculated with 1, 0 by-interated RES a - Day 0 challenged with 1, 0 or 100 plaque forming units 0 MA-EBOV intraperioneally set of MA-EBOV intraperioneally and the set of the set of the set of the set of the remain and real load, others observed up to 18 days victimis and real load, others observed up to 18 days - Lower EBOV doess repeated with 4 additional groups of 10 mice + KO mice followed protocol as above but challenged with r/SY/EBOV G on Day 0 instead of MA-EBOV 10 mice - both y y and Rec infection tested but Pic discontinued EX vitor experiments Macrophages challenged with serum from P. cynomolgi- inficient thesis macaques followed by r/SV/EBOV GP exposure Outcomes mortality, viternia and viral load (spleen, liver, kidneys)	•P-infected mice challenged with a low, but leftal. 1-in MA-EBOV dose showed reduced morbidity and mortality (P = 0.0076). These mice had 3-bg reduction in viral lotteness. International morbidity and mortality (P = 0.0076). These mice had 3-bg reduction in viral lotteness. International morbidity and mortality (P = 0.0076). These mice had 3-bg reduction and low-reduction faitherss. • • • • • • • • • • • • • • • • • • •
Clinical case r	eports				
	Study #	Publication	Locations	Patient details & methods	Relevant outcomes/findings
	21	Muehlenbachs et al. 2017. 215 (1):64-69.	Patient 1 from Gulu, Uganda Patient 2 from Isiro, DRC	Two pregnant EVD(+) patients calu, UBC in 2012 (BDB Writus) I sito, DBC in 2012 (BDB Writus) EBOV diagnostic 2 (BDB Writus) EBOV diagnostic 2 (BDB Writus) PBON RT-PCX & ELISA in Gulu • DDBV RT-PCX in isito Placenta, forati tissue and post-mortem skin biopsies performed and processed using standard histological methods	Patient 1: Distories with Sudan virus Outcone: why Sudan virus Outcone: a pointancus still birth, mother recovered Analysis of placenta: Analysis of placenta: Analysis of placenta: - immunohistochemical analysis showed EBOV antigen in placenta, primarily within areas of fibrin deposition, localised to maternal monomuclear cells including - immunohistochemical analysis showed EBOV antigen in placenta, primarily within areas of fibrin deposition, localised to maternal monomuclear cells including - immunohistochemical analysis showed EBOV antigen in placenta, primarily within areas of fibrin deposition, localised to maternal monomuclear cells including tarical is a fibrit to a struct a str
*Two pul demograj	blicati phic iı	ions from Haı nfo.	rtley et al. in relation to sam	10 cohort—PLoS Negl Trop Dis 11(2)e	005355 is source of prevalence and CFR data, PLoS Negl Trop Dis 11(2):e0005356 provides extra

https://doi.org/10.1371/journal.pone.0251101.t001

Table 1. (Continued)

investigating *Zaire ebolavirus* (ZEBOV, n = 12); only one investigated *Bundibugyo ebolavirus* (BDBV).

Malaria diagnosis was conducted by rapid diagnostic test (RDT) in most of these studies except two in which malaria diagnosis was made by qRT-PCR, with $Ct \leq 30$ indicative of a positive *Plasmodium* infection considered to provide equivalent sensitivity to microscopy [37, 38]. These two studies did, however, have overlap of the study population and only one explored the effect of parasite load on mortality-related outcomes using Ct level as a proxy [38]. Another two studies identified RNA reads mapping to *Plasmodium* to determine malaria infection prevalence.

Plasmodium falciparum (Pf) was specified as the causal agent of malaria in one cohort study [47] and one other estimated 95% of malaria infections in their sample were *Pf* although subsequent analyses included all species [37]. The mNGS studies also identified *Pf* as the causal agent in their samples. The rest did not specify Plasmodium species in their analyses but all used RDTs that detect all four major Plasmodium species (Pf, *Plasmodium vivax* (Pv), *Plasmodium malariae* (Pm) and *Plasmodium ovale* (Po)), except one which used a HRP2 RDT detecting Pf only [34].

Only one study [39] attempted to look at signalling/response pathways between EBOV (+)/Pf(+) and EBOV(+)/Pf(-) patients in an attempt to discern any immune response modulation, and one other compared clinical symptoms between the two infection groups [43].

Cross-sectional studies. One cross-sectional national survey was identified in Gabon which covered a general rural population of 4,272 and measured past exposure to ZEBOV infection using seroprevalence of ZEBOV-specific IgG antibodies, and current Plasmodium spp. infection using in-field blood smear and Plasmodium mitochondrial DNA sequencing. DNA sequencing identified Pf, Pm and Po infections within the sample but main analyses included all-species Plasmodium but was also reported for Pf and Pm infections only.

Clinical case reports. One clinical case report was identified describing the pathology of EVD and malaria coinfection during pregnancy. The clinical case report included one pregnant woman from Gulu, Uganda admitted with Sudan virus in 2000, and a second from Isiro, DRC, infected with BDBV in 2012. EVD was diagnosed using species-specific RT-PCR in each location as well as ELISA in Gulu. Histological methods were employed to analyse EVD and malaria pathology in placental and foetal tissues following delivery.

Laboratory studies. Both laboratory studies used an in vivo murine model of coinfection to analyse the impact of prior *Plasmodium* infection on outcomes related to EVD. The model in one study was of CD1 mice infected via intraperitoneal inoculation with *Plasmodium yoeli* (Py) at a dose of 10⁴ parasitized erythrocytes, followed by challenge with mouse-attenuated EBOV (MA-EBOV) at a 100 median lethal dose at different time points post Py infection. In each time-group, 6 mice were followed for survival to 28 days post infection (dpi), and four were euthanised on day 4-dpi to analyse blood and liver RNA concentrations for Py and MA-EBOV [49].

The second laboratory study used wild-type, interferon (IFN- $\alpha\beta$) receptor knock-out mice (Ifnar⁻¹⁻), and IFN- γ receptor null mice (Ifngr⁻¹⁻) inoculated intraperitoneally with a higher dose of 10⁶ parasitized red blood cells of Py and challenged with MA-EBOV or recombinant vesicular stomatitis virus (rVSV)/EBOV glycoprotein (GP) at three different concentrations: 1-, 10- and 100-iu on the sixth day following Py infection. In each group, seven mice were followed for survival up to 18 days and three were euthanised on day 3 to determine viremia and viral load. Knock-out mice were used to study the signalling pathways implicated in initial lab findings. The study also analysed Ifnar⁻¹⁻ mice with *P. chabaudi* (Pcc) infection in the first round of experiments but found no difference in outcomes compared to Py so was discontinued. To test the durability of *in vivo* effects, parallel experiments challenged Py-infected mice

with rVSV/EBOV GP at weeks 1, 3, 5, and 7 post-Py inoculation. In addition, an *ex vivo* model of infection was analysed using naïve human and mouse macrophages challenged with sera from parasitaemic rodents and from rhesus macaques infected with *Plasmodium cynomolgi* [50].

Review of key findings

Prevalence and seroprevalence of EBOV/*Plasmodium* **spp. coinfection**. Thirteen cohort studies reported data on prevalence of coinfection. Prevalence of *Plasmodium* coinfection among EVD patients was found to be 33.3% among patients with BDBV in DRC though this was only based on nine malaria RDTs. Among patients infected with ZEBOV, prevalence of *Plasmodium* coinfection in EVD(+) cohorts ranged from 18.9% to 72.2% for studies utilising malaria RDTs, and 15.4% and 19.4% for the two studies utilising qRT-PCR. In addition to these cohorts, RNA sequencing data from EBOV(+) cases in Guinea, found that 84% (156/186) contained reads mapping to Pf, higher than malaria prevalence found by RDT on the same sample of patients (33%, 40/121). However, read depth, a proxy for parasitaemia density, varied widely from 59 to 358820 FPKM (a measure for read depth) and the discrepancy between diagnostic methods reduced as number of reads mapping to *Plasmodium* increased. RNA sequencing in Boende, DRC, found 24.3% of EBOV(+) cases were coinfected with Pf. One study which focused only on children aged <18years found prevalence in this group to be 40%. Prevalence of *Plasmodium* infection was reported as higher in children aged and less common in EBOV(+) compared to EBOV(-) individuals [34, 46].

The cross-sectional survey found evidence of an epidemiological overlap of the two infections with 10.2% (n = 425) of adults in rural Guinea positive for both current *Plasmodium* infection and ZEBOV-specific IgG antibodies, a significant overabundance than would be expected by chance (χ^2 = 59.4, df = 1, p<0.0001).

Eight observational cohorts were included in the meta-analysis. The prevalence of *Plasmo-dium* coinfection among EVD patients varied from 18.9% to 44.4% between studies, with the overall meta-analysis estimate calculated at 21.7%, 95% CI 18.7–25.1% (Fig 2). Between-study heterogeneity was low-to-moderate with $I^2 = 36\%$, 95% CI 0–72%; chi² = 11.01, p<0.14; and Tau² = 0.05. The prediction interval was 13.3–33.4%. Sub-group analysis including only studies with ZEBOV as the causal agent had a similar overall estimate and similar heterogeneity. Sub-group analysis by malaria diagnostic showed a similar overall estimate and high heterogeneity from studies utilising only RDTs compared to qRT-PCR or NGS an RT-PCR, though these latter methods only included one study each (S1 and S2 Figs).

Effect on disease pathology and the immune response. *Human subjects*. Description of the impact of *Plasmodium* coinfection on EVD pathology and immune response was given in the clinical case study and in both metagenomic studies. The clinical case report presented evidence of a potential pathophysiological interaction of the two infectious agents in the placenta [51]. Although she recovered from infection, one pregnant woman infected with Sudan virus in Uganda had a spontaneous still birth during her illness. Subsequent histological and immunohistochemical analysis showed colocalization of malarial parasite pigment and EBOV antigen within the placenta. Malarial parasite pigment was observed in fibrin and within macrophages embedded within the fibrin. EBOV antigen was similarly primarily localised to areas of fibrin deposition and maternal mononuclear cells, including the malarial parasite-pigment-laden macrophages. This interaction was not observed in a second pregnant woman infected with BDBV in DRC despite testing positive for malaria by RDT. She was, however, kept on a course of artemether-lumefantrine following the positive RDT result. Information on malaria diagnosis and treatment was not reported from the clinical case in Uganda.

First author	# Plasmodium(+)	# Malaria tests	Weight	% Prev [95% C	1	:				
Kratz	4	9	2.3%	44.4 [13.7; 78.8]						
Overall		9	2.3%	44.4 [17.7; 74.9]					-	
Heterogeneity: n	ot applicable									
EBOV Species	s = ZEBOV									
Gignoux	65	337	16.1%	19.3 [15.2; 23.9]		-				
Rosenke	185	956	19.5%	19.4 [16.9; 22.0]		-+-				
Kerber	261	1091	20.1%	23.9 [21.4; 26.6]		-+				
Hartley	35	145	12.8%	24.1 [17.4; 31.9]		-	_			
Waxman	53	254	15.1%	20.9 [16.0; 26.4]		-				
Vernet	14	74	8.2%	18.9 [10.7; 29.7]		-	-			
Li	9	37	5.8%	24.3 [11.8; 41.2]						
Overall		2894	97.7%	21.4 [19.3; 23.6]		•				
Heterogeneity: T	au ² = 0.0087; Chi ² =	8.49, df = 6 (P = 0	.20); $ ^2 = 2$	29% [0%; 70%]						
Overall		2903	100.0%	21.7 [18.7: 25.1]	1	•				
Prediction inte	erval			[13.3: 33.4]		_	_			
Heterogeneity: T	au ² = 0.0526; Chi ² =	11.01, df = 7 (P =	0.14); I ² =	36% [0%; 72%]		1		I		
					0	20	40	60	80	100
							% Prev	valence		

Fig 2. Meta-analysis of the prevalence of Plasmodium infection among EVD(+) cases, overall and split by species of Ebolavirus.

https://doi.org/10.1371/journal.pone.0251101.g002

In their metagenomic sequencing study, Li et al. found no significant differences in clinical characteristics (symptoms including, for example, fever, fatigue, bleeding and pain) between EVD(+)/Pf(+) and EVD(+)/Pf(-) patients [43]. Differences were observed by Carroll et al.'s comparison of biomarkers of the immune response between EBOV(+)/Pf(+) and EBOV(+)/Pf(-) patients, albeit only a small sample of each (n = 10 and 13, respectively) [39]. The abundance of transcripts of genes related to the innate response to severe malaria infection (IFIT2, IFIT3, IFITM3, ISG15, STAT1, MX1, TGF- β , PRF1, PGRMC1, CTSW, ICAM-1, CD36, IFN- γ , interleukin-10, and IFN regulatory factor 9) were not significantly different between the two groups, suggesting *Plasmodium* infection did not have any additional effect on the immune response to EBOV infection. However, biomarkers associated with the coagulation pathway (FGA, FGB, FGG, FGL1, and ALB) were increased in abundance in EBOV(+)/Pf(-) cases, suggesting, vice versa, that Pf coinfection had an inhibitory effect on coagulation.

Animal models. One experimental lab study measured immune response biomarkers in infected mice and found that acute *Plasmodium* infection protected against a low but lethal dose (1-iu) of virus (both MA-EBOV and rVSV/EBOV) by eliciting the proinflammatory interferon gamma (IFN- γ) pathway, rendering cells resistant to EBOV infection [50]. Morbidity and mortality were reduced, as well as viremia and viral load in the liver, spleen and kidneys of coinfected mice. The same protection was not conferred by administration of a single large bolus of recombinant IFN- γ , suggesting a persistent, subpatent *Plasmodium* infection and/or a sustained level of IFN- γ confers protection against EBOV viral replication (effect on mortality discussed below).

Effect on mortality. *Human subjects*—*CFR*. Crude EVD-related mortality was reported in 12 cohort studies and ranged from 44% (n = 39/90, ZEBOV infection, Guinea) to 72.8% (n = 118/162, ZEBOV infection, Guinea). For BDBV infection, crude CFR was 53.8% (n = 28/ 52, DRC) and in the study looking only at children aged <18 years crude CFR was 56.6% (n = 69/122, Liberia & Sierra Leone). CFR by *Plasmodium* infection status were reported in eleven cohorts and ranged from 42.2% to 85.7% for EBOV(+)/Pl(+) cases, and from 52.2% to 60.3% for EBOV(+)/Pl.(-) cases. Eight of the cohort studies were included in a meta-analysis for CFR estimates (Fig 3). Overall crude CFR was found to be 57.6%, 95% 54.0–61.1% and between-study heterogeneity was moderate-to-high with I^2 at 68%, 95% CI 33–85%, $Chi^2 = 21931$, p<0.01 and $Tau^2 = 0.02$. The prediction interval was 48.7–66.0%.

CFRs by malaria infection status were estimated at 62.1%, 95% CI 50.1–72.8% for EBOV (+)/Pl.(+) cases and 56.7%, 95% CI 53.6–59.7% for EBOV(+)/Pl(-) cases (Fig 3). Heterogeneity between studies was high for the measure of CFR among coinfected cases ($I^2 = 75\%$, 95% CI 49–88%, Chi² = 27.76, p<0.01, Tau² = 0.2) but much lower for the CFR of singly-infected cases ($I^2 = 5\%$, 95% CI 0–69%, Chi² = 7.33, p = 0.39, Tau² = 0.01).

Each CFR was similar when looking only at ZEBOV species alone (Fig 3) and when looking at studies that used malaria RDTs only (S1 and S2 Figs). Although heterogeneity between study estimates of CFR among coinfected cases was reduced when analysing only studies that used malaria RDTs (S1 and S2 Figs).

Human subjects—effect estimates. Seven studies found no significant difference in CFR by *Plasmodium* infection status in unadjusted analyses [36, 39, 40, 42, 43, 45, 47]. Within these, Kratz et al., Vernet et al and Li et al. had a small sample sizes of coinfected individuals (n = 3, 14 and 7, respectively) and Garbern et al. had a high amount of missing malaria diagnostic data. Carroll et al. reported that mortality appeared to be increased by Plasmodium coinfection using RDT data, but this was not found using prevalence as determined by genomic sequencing data. They did find that higher burden of Plasmodium infection appeared to have higher mortality, but no absolute correlation was observed.

Two studies found no effect of coinfected in adjusted analyses. Hartley et al. found coinfection was associated with significantly higher mortality when adjusted only for age and sex (53.6% v 74.3%, OR = 3.9, p = 0.03), however, these individuals also had higher viral loads and once this was taken into account the detrimental effect was abrogated. Gignoux et al. found no effect after adjusting for multiple confounders including antimalarial prescription, age group, sex, Ct value at admission, days from symptom onset to admission, receipt of IV fluids, and number of inpatients at the Ebola treatment centre on day of admission (aRR = 1.23, 95% CI 0.52–2.86). However, this study did find that *Plasmodium* coinfection inhibited the apparent beneficial effect of ASAQ treatment; ASAQ treatment appeared to reduce mortality in EBOV (+)/Pl(-) cases compared to those treated with A-L whereas no beneficial effect was seen in EBOV(+)/Pl(+) cases.

Two studies reported increased mortality in either the general study population (Waxman et al.) or in association with age (Kerber et al.) In the former, although no significant effect was found from a smaller study drawn from the same source population (Garbern et al.), with a larger sample Waxman et al., found a significant detrimental effect of *Plasmodium* coinfection on mortality with an elevated CFR (52% v 66%) and 28-day mortality (8.2 per 1,00 persondays). In a focused subgroup analysis, adjusted for age only, there was greater mortality among EBOV(+)/Pl(+) compared to EBOV(+)/Pl.(-) cases (aHR 1.69, 95% CI 1.14–2.52, p = 0.009). Compared to EBOV(-)/Pl(-) patients, the hazard of dying was highest in EBOV(+)/Pl.(+) cases (aHR 9.4, 95% CI 6.2–14.2, p < .0001) followed by EBOV(+)/Pl.(-) cases (aHR 6.0, 95% CI 4.4–8.0, p < .0001). Kerber et al, meanwhile, found that *Plasmodium* coinfection only had an impact on mortality among 5–15-year-olds among whom it increased CFR by >20% and increased odds of death to 4.2 times greater than individuals of the same age group without *Plasmodium* co-infection. No other age groups appeared to be affected, suggesting an interaction between age and infection status.

In contrast, just one study found coinfected patients had significantly lower CFR than singly infected cases (p = 0.007) [38]. *Plasmodium* infection was associated with an increased survival when adjusted for age, sex and EBOV viral load (aRR = 1.2, 95% CI 1.1–1.4, p = 0.004).



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	EBO	/(+)/PI.(+)	EBO\	/(+)/PI.(-)					
First author	# Deaths	# Cases	# Deaths	# Cases	Weight	RR [95% CI]		3 .	
Gignoux	34	56	144	239	15.1%	1.01 [0.80; 1.27]			
Rosenke	78	185	416	771	17.5%	0.78 [0.65; 0.94]			
Kerber	150	249	452	798	20.3%	1.06 [0.95; 1.20]		-	
Hartley	26	35	70	110	14.8%	1.17 [0.92; 1.49]			
Waxman	35	53	105	201	15.1%	1.26 [1.00; 1.60]			
Vernet	12	14	36	60	12.4%	1.43 [1.06; 1.92]			
Li	5	7	10	18	4.8%	1.29 [0.69; 2.40]			
Overall		599		2197	100.0%	1.09 [0.90; 1.31]		-	
Prediction inte	erval				120	[0.68; 1.74]			
Heterogeneity: T	au ² = 0.0271	1; Chi ² = 18	3.09, df = 6	(P < 0.01);	$l^2 = 67\%$	[26%; 85%]	I		I
							0.5	1	2
								Risk Ratio	

Fig 4. Meta-analysis of unadjusted risk ratio estimates for the effect of Plasmodium infection on EVD-related mortality (all ZEBOV).

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However, the effect was more pronounced when stratified by plasmodium parasitaemia level, using Ct as a proxy. High Plasmodium burden was then associated with increased risk of survival to 1.4, 95% CI 1.2–1.7, p<0.0001) whereas low parasite burden infection did not have a significant effect (RR 1.2, 95% CI 0.98-1.4, p = 0.08) P = 0.007). These effect estimates did not include individuals aged <5 years due to low numbers of cases in that age group.

In addition to *Plasmodium* infection, studies also reported other significant variables affecting CFR as including age, viral load, aspects of clinical treatment (e.g. referral time, length of stay) and certain clinical features (e.g. bleeding and other symptoms).

For the meta-analysis, numbers of exposed/unexposed and with outcome data was extracted from seven cohort studies, all investigating ZEBOV (Fig 4). Individual (unadjusted) RRs ranged from 0.78, 95% CI 0.65-0.94, to 1.43, 95% CI 1.06-1.92. The overall unadjusted RR was estimated at 1.09, 95% CI 0.90–1.31. There was moderate-to-high heterogeneity ($I^2 =$ 67%, 95% CI 26–85%, Chi² = 18.09, p<0.01, Tau² = 0.03) and a prediction interval of 0.68– 1.74.

Since Rosenke et al. and Li et al. used different and more sensitive diagnostic methods for malaria detection (qRT-PCR and mNGS), these were excluded and the meta-analysis re-run only on the studies using RDT. This estimated a higher but still insignificant risk ratio of 1.15, 95% CI 0.98–1.35, with lower heterogeneity ($I^2 = 24\%$, 95% CI 0–69%, $Chi^2 = 5.24$, p = 0.26, $Tau^2 = 0.009$) and slightly reduced prediction interval of 0.80–1.64 (Fig 5).

Only three studies reported final adjusted effect estimates for the whole study population, though with varying confounders adjusted for. These three estimates pooled to a combined

	EBOV(+)/PI.(+)	EBOV(·	+)/Pl.(-)			
First author	# Deaths #	Cases #	Deaths #	Cases	Weight	RR [95% CI]	
Gignoux	34	56	144	239	18.1%	1.01 [0.80; 1.27]	
Kerber	150	249	452	798	33.0%	1.06 [0.95; 1.20]	
Hartley	26	35	70	110	17.5%	1.17 [0.92; 1.49]	
Waxman	35	53	105	201	18.1%	1.26 [1.00; 1.60]	
Vernet	12	14	36	60	13.2%	1.43 [1.06; 1.92]	
Overall		407		1408	100.0%	1.15 [0.98; 1.35]	
Prediction inte Heterogeneity: 7	erval Fau ² = 0.0094; (Chi ² = 5.24	l. df = 4 (P =	= 0.26); 1	² = 24% [0	[0.80; 1.64] 0%: 69%]	
				,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,			0.75



Fig 5. Meta-analysis of unadjusted risk ratio estimates for the effect of Plasmodium infection on EVD-related mortality (all ZEBOV) including only studies with malaria RDT as the diagnostic.

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Fig 6. Meta-analysis of adjusted risk ratio estimates. A) all adjusted RR estimates, and B) only those that included adjustment for viral load.

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aRR estimate of 1.14, 95% CI 0.41–3.17 with high heterogeneity ($I^2 = 82\%$, 95% CI 45–94%, $Chi^2 = 11.22$, p<0.01)) and an incomprehensibly wide prediction interval (Fig 6A). Since only two of these adjusted for EBOV viral load, which was considered to be a significant factor in predicting outcome in seven studies, a meta-analysis was conducted solely on these two studies, estimating a RR of 0.86, 95% CI 0.12–6.30 (Fig 6B).

Animal models. In addition to these observational studies in the field, the two laboratory studies also report the effect of coinfection on mortality in murine models. Rosenke et al. 2018 found no effect of coinfection on mortality in mice when challenged with a high dose of MA-EBOV at 100-median lethal dose. In contrast, Rogers et al. found that acute *Plasmodium* infection protected from lethal virus challenge at low (1-iu) MA_EBOV challenge but less so with moderate to high (10-iu and 100-iu) doses. This protective effect waned over time, with Py-infected mice protected from EBOV challenge three weeks after Py infection, but not by 5 weeks. This was IFN-γ dependent as discussed above.

Quality of studies included in meta-analysis. The eight studies included in the metaanalysis were analysed for quality using the Newcastle-Ottawa Scale (Table 2). Four studies (Kratz, Waxman, Vernet and Li) were deemed to be 'Poor' because, while they scored well under the selection and outcome categories, they did not score under comparability, i.e. they did not attempt to adjust analyses to take into account important confounding factors. Although the other four were deemed 'Good', all studies lost points on the representativeness of the exposed cohort since all only had diagnostic data from hospital-based cohorts, and four studies had follow-up rates that were deemed high enough to introduce potential bias into their final estimates (Kratz, Gignoux, Rosenke and Vernet).

Discussion

We have attempted to consolidate findings on the prevalence and effect of *Plasmodium* coinfection on mortality from EVD which could have significant implications for future outbreaks of EVD and other viral outbreaks with similar pathology. The number of studies identified, particularly those for the meta-analysis, was limited. This could be because EVD outbreaks have historically been small and, indeed, the majority of studies stem from the 2014 West

		Selection			Comparability		Outcome		Quality
Study	Representativeness of Exposed Cohort	Selection of the Non- Exposed Cohort from Same Source as Exposed Cohort	Ascertainment of Exposure	Outcome of interest not present at start of study	Comparability of cohorts based on factors controlled for	Assessment of outcome	Follow-up was long enough for outcome to occur	Adequacy of follow-up	Score (/9)
Kratz	Although the paper looked at community cases, malaria diagnostic data was only collected among cases that made it to the ETC which the paper reports as having different characteristics to the community cases	Yes *	Secure clinical data collected by trained medical staff *	Yes [#] *	CFR by malaria status were reported but no attempt to control for confounding factors	Record linkage *	Yes—cases followed until death or discharge from ETU *	Cases all followed for outcome but only 50% of ETC patients had malaria diagnostic test data available for reasons unknown.	5—Poor
Gignoux	ETC-based cohort in Foya, Liberia. Could be a biased subsample of community cases but no comparison of sample demographics to general population	Yes *	Secure clinical data collected by trained medical staff *	Yes [#] *	CFR/Effect measures adjusted for Ebola viral load. * Other factors adjusted for included age, sex (among others) but didn't measure malarial parasite load.	Record linkage *	Yes—cases followed until death or discharge from ETU *	Malaria diagnostic data missing for 44/ 381 EBOV(+) cases (11.5%). Outcome data missing for 53/381 EBOV(+) cases (13.9%).	6—Good
Rosenke	ETU-based cohort j Monrovia, Liberia. Could be biased subsample of community cases but no comparison of sample demographics to general population	Yes *	Secure clinical data collected by trained medical staff *	Yes [#] *	Outcome measures adjusted for Ebola viral load. * Outcome measures also stratified by malarial parasite load and adjusted for age and sex. *	Record linkage *	Yes—cases followed until death or discharge from ETU *	Malaria diagnostic data missing for 226/1182 EBOV(+) cases (19.1%).	7—Good
Kerber	Study looked at community deaths as well as ETU-based cases but only ETU cases had malaria diagnostic data available, thus could represent a biased sub-sample	Yes *	Secure clinical data collected by trained medical staff*	Yes [#] ∗	Outcome measures adjusted for Ebola viral load, but only viral load. *	Record linkage *	Yes—cases followed until death or discharge from ETU *	Only 2% (26/1231) EBOV(+) patients without outcome data; unlikely to introduce significant bias. *	7—Good
Hartley	ETC-based cohort in Sierra Leone. Could represent a biased sub-sample of all community cases if inequitable accessibility/ uptake in the community; no comparison made of sample demographics to general population	Yes *	Secure clinical data collected by trained medical staff *	Yes [#] *	Outcome measures adjusted for Ebola viral load, but only viral load.*	Independent blind assessment *	Yes—cases followed until death or discharge from ETU *	Only 13/158 EBOV (+) cases missing malaria diagnostic data (8.2%); unlikely to introduce significant bias. *	7—Good
Waxman	ETU-based cohort spread over 3 ETUs. Could represent biased sub-sample of all community cases; no comparison made of sample demographics to general population or deaths in the community.	Yes *	Secure clinical data collected by trained medical staff *	Yes [#] *	Outcome measures adjusted for age only.	Record linkage *	Yes—cases followed until death or discharge from ETU *	Complete *	6—Poor
Vernet	ETC-based cohort in Macenta, Guinea. Could represent biased sub-sample of all community cases; no comparison made of sample demographics to general population or deaths in the community.	Yes *	Secure clinical data collected by trained medical staff *	Yes [#] *	Outcome measures not adjusted for any confounding variable	Record linkage *	Yes—cases followed until death or discharge from ETU *	12.5% non-fatal and 26.3% fatal EVD cases without clinical follow-up, and 24% cases without data on viral titre or malaria diamosis	5—Poor

Table 2. Outcome of Newcastle-Ottawa Scale analysis on studies included in the meta-analysis.

(Continued)

Table 2. (Continued)

		Selection			Comparability		Outcome		Quality
Study	Representativeness of Exposed Cohort	Selection of the Non- Exposed Cohort from Same Source as Exposed Cohort	Ascertainment of Exposure	Outcome of interest not present at start of study	Comparability of cohorts based on factors controlled for	Assessment of outcome	Follow-up was long enough for outcome to occur	Adequacy of follow-up	Score (/9)
Li	ETU-based cohort in Boende, DRC. Could represent biased sub-sample of all community cases; no comparison made of sample demographics to general population or deaths in the community.	Yes *	Secure clinical data collected by trained medical staff *	Yes [#] ∗	Outcome measures not adjusted for any confounding variable	Record linkage *	Yes—cases followed until death or discharge from ETU *	5/70 patients without clinical data (7%); unlikely to introduce significant effect on results. *	6—Poor

[#]Given the acute nature of Ebola outbreaks, all studies were deemed to not have the outcome of interest prior to the study start.

Comparability stars awarded if study adjusted for Ebola viral load (one star) and/or adjusted for age, sex and Plasmodium parasiteamia (one star). Assessment of quality scale =

Good quality: 3 or 4 stars in selection domain AND 1 or 2 stars in comparability domain AND 2or 3 stars in outcome/exposure domain.

Fair quality: 2 stars in selection domain AND 1 or 2 stars in comparability domain AND 2 or 3 stars in outcome/exposure domain.

Poor quality: 0 or 1 star in selection domain OR 0 stars in comparability domain OR 0 or 1 stars in outcome/exposure domain.

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Africa EVD outbreak which had a huge affected population in comparison and a high level of support from national and international NGOs and research institutions [6, 52]. EVD outbreaks also develop quickly and in countries with limited surveillance and weak health systems, limiting resources and planning for research purposes. Although we identified 13 observational studies, there were overlapping study populations meaning only eight unique studies could be included in the meta-analyses. This is largely because research data came from ETUs which were set-up to specifically manage EVD patients during the 2014 outbreak.

Overall, one-quarter of EVD cases were infected with *Plasmodium* (26.9%, 95% CI 16.8–40.0%), and, aside from one, appeared relatively consistent across individual studies. Plasmodium spp. prevalence was lower in EVD confirmed cases compared to patients without EBOV infection, most likely because presenting symptoms were due to the viral infection and not malaria in the former group. Indeed, Hartley et al. found that malaria infection could be used as one variable (among others) to help distinguish EVD cases from other diagnoses [35]. Despite this, one-fifth of EVD cases is a high proportion and would have important public health implications if coinfection were to alter morbidity or mortality in a significant way. There was a lack of consensus, however, between studies in relation to coinfection and its effect on EVD outcomes.

From the cohort studies reviewed, the majority found no significant effect of *Plasmodium* infection on mortality, while two found a significant detrimental effect, and one found a significant protective effect. Overall, the meta-analysis estimated that the CFR of coinfected patients was not significantly different from the CFR of EBOV(+)/Pl(-) patients, and a meta-analysis of effect sizes found no significant effect of coinfection on mortality (RR = 1.09, 95% CI 0.90– 1.31). Although tests of heterogeneity have low statistical power with a small number of studies, use of the chi-square p-value and I² confidence intervals can provide more confidence in these estimates and in this case suggest high heterogeneity between studies for both the overall effect estimate and the CFR of coinfected patients (EBOV(+)/Pl(+)) [53]. Conversely, between-study heterogeneity appeared to be lower between estimates of the CFRs of individuals infected with EBOV alone (EBOV(+)/Pl(-)). The observed heterogeneity in the estimates

related to coinfection could suggest other factors complicate the outcome that have not been accounted for.

In terms of study protocols, only two studies included in the meta-analysis used a malaria diagnostic that was more sensitive than RDT: qRT-PCR and a combination of RT-PCR and mNGS. One of these [38] (using RT-PCR) was also the only one to find a positive effect of coinfection on surviving EVD. When these studies were removed from the meta-analysis, heterogeneity was reduced and the overall estimate, although it remained insignificant, was biased toward a detrimental effect of *Plasmodium* on EVD mortality. However, the difference in testing procedure doesn't logically explain why Rosenke et al. would observe a protective effect from *Plasmodium* since RT-PCR is more sensitive than RDT, particularly in detecting low parasite density infection, and so could be expected to identify more malaria cases and would actually bias the effect estimate toward a detrimental effect (a higher RR if considering mortal-ity as the outcome) [54, 55]. Conversely, the studies using RDTs could be biased toward a detrimental effect since they used RDTs based on HPR2 for Pf identification which has been shown to be associated with false positive results [56, 57].

Additionally, the RT-PCR and mNGS tests could detect different stages of malaria infection to the RDTs, i.e. detecting sub-patent infection compared to more active/acute infection. These stages of malaria are associated with different immune responses which could affect EVD outcomes differently. Younger age groups are more at risk of acute *Plasmodium* infection compared to adults who gain some level of immunity to severe infection and are more likely to have a chronic, subpatent infection [58, 59]. In the cohort studies presented here, younger people were found to be more likely to be associated with both coinfection and greater EVD-related CFR yet age was not adjusted for in most analyses. If a study had a younger age profile this could result in an inflated CFR among coinfected cases. Furthermore, Rosenke et al.'s analysis did not include cases age <5 years, while other evidence pointed to a potential interaction between age and Plasmodium infection status; Kerber et al. found that coinfection among 5-14-year olds increased their risk of death but the same was not seen in other age groups [46]. This could be suggestive of differences in the intensities of infection and/or immune responses between age groups in either EBOV or *Plasmodium* infection.

Acute Plasmodium infection is associated with activation of pro-inflammatory pathways and has been implicated in influencing response to other concurrent infectious agents such as respiratory viruses, enteric bacteria and HIV [17-20]. HIV, like EBOV, is an RNA virus, however, it is a chronic infection as opposed to the acute infection seen in EVD which could lead to alternative interactions. However, prior Plasmodium infection was found to reduce viremia and associated pathologies of chikungunya virus, a more acute RNA virus, via proinflammatory responses through IFN- γ production [60]. Despite this, only one study in our meta-analysis attempted to discern the effect of more acute Plasmodium infection. Rosenke et al. looked at parasite burden (using Ct as a proxy) and found that higher parasite load (Ct \leq 20), which would be expected from more acute infection, was associated with improved survival (reduced mortality) compared to those with a lower parasite burden for which protection was not significant. A similar protective effect from acute infection was seen in the laboratory study by Rogers et al which found acute Plasmodium infection protected mice and human macrophages from EBOV challenge via a low but sustained IFN-γ response which protected from EBOV challenge up to three weeks post *Plasmodium*-infection but dwindled thereafter. However, in the only study to monitor similar human biomarkers, Carroll et al found that markers of the innate immune response to acute *Plasmodium* infection, including IFN- γ , were not different between Pf(-) and Pf(+) EVD cases whereas biomarkers associated with the coagulation pathway were reduced in patients Pf(+). This analysis thus suggests the inflammatory pathway

found to be important in mice is perhaps not important in human cases, although, the analysis by Carroll et al was based on a very small sample size of individuals (10 and 13 per group).

The protective effect observed from the laboratory study by Rogers et al. was not observed in the second laboratory study identified, however, this second study used only a very high lethal dose of MA-EBOV (100-iu) [49]. Use of similarly high doses by Rogers et al. (10-iu and 100-iu) also showed limited effect on morbidity and outcomes compared to the low EBOV dose (1-iu) which was significantly protective, thus suggesting high viral load overrides any potential protection from coinfection [50]. In human studies, viral load was only quantified in seven studies and, where analysed, all found that higher viral load led to greater mortality. Despite this, only three studies reported analyses that adjusted for viral load. Among these, Hartley et al. found that coinfected cases had a higher viral load than EBOV-only infected cases and when accounted for in their analyses viral load abrogated the apparent detrimental effect of *Plasmodium* infection on mortality [34]. The two other studies that adjusted for viral load included the positive effect estimate from Rosenke et al. and an insignificant finding from Gignoux et al. [36, 38]. Not accounting for this confounding variable is a clear limitation in the other effect estimates reported.

Due to the failure to adjust for important confounders, four of the eight studies included in the meta-analysis were deemed poor quality based on the Newcastle-Ottawa Scale analysis. As well as not adjusting for viral or parasite load, many of the studies failed to account for other potential confounders including age, sex, and time from symptom onset to admission. The majority of studies did not specify *Plasmodium* species and it could be expected that different species would be associated with a different proportion of chronic versus acute infections and differences in immune response. However, it is unlikely to explain differences in this context since the vast majority of malaria infection is caused by Pf in each of these countries [61]. Similarly, it is conceivable that different EBOV species could have different immune responses that may interact differently. However, all but one observational study examined ZEBOV and thus does not explain differences observed.

Outside of the cohort and laboratory studies, the case report of a pregnant woman in Uganda implicated other potential interactions between the two infectious agents in the placenta of pregnant women [51]. It is unclear whether the co-localization of these two pathogenic agents had an impact on the outcome of still birth experienced in this case. As a one-off observation, this does need to be explored in other future cases to see if there is a legitimate interaction of concern.

So, what does all this mean and what are the implications for future outbreaks of EVD and other viral diseases? From the literature as it stands, it remains unclear whether *Plasmodium* coinfection has an effect on EVD morbidity and mortality. Although pooled meta-analyses suggest no significant effect, there is a high amount of variability between study effect estimates that have not all had appropriate confounding variables considered, and/or had follow-up rates that may have biased results. With the advent of the new Ebola vaccine (rVSV-ZE-BOV-GP, manufactured by Merck) which has shown high efficacy in trials, the clinical interaction of comorbidities with EBOV may have less significance in future EVD outbreaks [21]. However, it will take time for a vaccine to be delivered widely and is not yet fully assessed in all populations. Furthermore, the learnings described here in terms of research conducted and gaps remaining can be applied to improve our knowledge of malaria coinfections on other viral outbreaks in the future. Since other viral diseases with similar immune response profiles could be affected similarly by Plasmodium infection, improvements in study design and data collection could be made to ensure that all essential clinical information is collected during subsequent outbreaks. In this case, features of clinical presentation, details of clinical acute

care, data on viral loads, parasite densities and immune system biomarkers in particular are important and lacking.

Planning for research purposes is difficult in an outbreak setting where the outbreak is fastmoving and resources are limited but being better prepared with a planned set of guidance on what data should be collected prior to the next outbreak would enable faster implementation of data collection for subsequent research. Following analyses, all studies should be encouraged to publish all results related to these important factors, including raw numbers, unadjusted and adjusted estimates, to better enable meta-analytics.

Even despite these improvements, due to the complex nature of the immune system, it may be that experimental lab methods will give the best insights. Findings by Rogers et al are interesting, but more comparison needs to be done to look into other disease models, as well as how well these animal models transfer to the human context. Mouse models of malaria and EVD do not mimic all facets of the diseases in humans, but findings from these models can be used to narrow down the field of investigation in the clinical setting—e.g. explicit measurement of immune system biomarkers such as IFN- γ , stratified by Plasmodium parasite load and viral load, should now be recommended in future clinical research.

Remaining limitations on implementing better-planned research may be sample size restrictions if future outbreaks are not large enough, and diagnostic and clinical capacities to conduct the research. In this case, retrospective genomics analysis on stored blood samples may offer a useful tool since this is increasingly affordable and samples can be stored and analysed in retrospect or in partner institutions with greater research capacity. RNA/DNA read data can provide quantifiable information on prevalence of infection, viral and parasite loads, and biomarkers in one protocol, and thus perhaps be able to answer some of gaps in the data presented here.

Supporting information

S1 Checklist. (DOC)

S1 Fig. Prevalence of Plasmodium coinfection among EVD cases by malaria diagnostic. (DOCX)

S2 Fig. CFR of EVD cases by Plasmodium infection including only studies using RDT for malaria diagnosis.

(DOCX)

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Author Contributions

Conceptualization: Hannah M. Edwards, James K. Tibenderana.

Data curation: Hannah M. Edwards, Helen Counihan, James K. Tibenderana.

Formal analysis: Hannah M. Edwards, Helen Counihan, Craig Bonnington, Jane Achan, Prudence Hamade, James K. Tibenderana.

Methodology: Hannah M. Edwards, James K. Tibenderana.

Validation: Hannah M. Edwards.

Visualization: Hannah M. Edwards.

Writing - original draft: Hannah M. Edwards, James K. Tibenderana.

Writing – review & editing: Hannah M. Edwards, Helen Counihan, Craig Bonnington, Jane Achan, Prudence Hamade, James K. Tibenderana.

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